Effect of temperature on stable H and O isotope composition of a freshwater bacterioplankton

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Microbial communities play a crucial role in biogeochemical processes, yet their responses to environmental conditions remain complex and poorly constrained. Stable isotope fractionation serves as a powerful tool for investigating biogeochemical cycling and ecological changes by providing insights into environmental stressors, such as climate change, as has been demonstrated by the temperature dependent fractionation of ^{2/1}H incorporated into algal lipids [1]. Until now, there is little evidence extending such isotope systematics to the microbial loop. In this study, we investigated the growth signals of Limnohabitans, a cosmopolitan freshwater pelagic prokaryote, under controlled temperature conditions (8, 16, and 24 °C) in deuterium- and ¹⁸O-enriched media to track isotope water incorporation into biomass. Distinct isotopic fractionation patterns were observed for biomass incorporation of water hydrogen (-43.5 > $^{2}\epsilon$ > -12.3%) and oxygen (23.6 > $^{18}\epsilon$ > 27.3‰) and were higher in magnitude at the optimal growth temperature (16°C). These ² \varepsilon values are lower than reported for algal lipids [1], which may, in part, be explained by the incorporation of H from organic sources into Limnohabitans biomass. This notion is supported by relatively low water-H assimilation factor (a_w) determined for the bacterioplankton (0.3-0.5), which were temperature dependent and possibly related to metabolic demands. This study additionally reports among the first 18 values for bacterioplankton, which were consistently positive and similar to that reported for cellulose and other sugars formed in leaves. Our findings highlight the importance of cultivation techniques for improving the investigation of stable isotope signals in the environment and the role of microbial communities in aquatic ecosystems. Further understanding of stable H and O isotope fractionation by bacterioplankton can provide deeper insights into their metabolic demand and growth and thereby offer a new tool to estimate microbial contributions to aquatic C cycling.

[1] Sachse, D., et al. (2012). Molecular paleohydrology: Interpreting the hydrogen-isotopic composition of lipid biomarkers from photosynthesizing organisms. Annual Review of Earth and Planetary Sciences, 40(1), 221–249. https://doi.org/10.1146/annurev-earth-042711-105535

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