

Modeling stable isotope fractionation in microbial methane production

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Methanogenic archaea produce methane (CH₄) from CO₂ and H₂ or from organic matter in anoxic environments. This process is associated with characteristic isotopic fractionations of C and H isotopes between the produced methane and its substrates (denoted by ¹³α and ²α for singly substituted isotopes, and Δ₁₈ for clumped isotopologues of CH₄). Accordingly, the isotopic composition of CH₄ is used to trace its sources in the environment. The isotopic fractionation during methanogenesis is generally correlated with the net thermodynamic drive of the methanogenic reaction (ΔG_r) [1–3]. However, there is no mechanistic explanation for larger-than-equilibrium fractionations of C isotopes at relatively small ΔG_r, and for the weaker dependence of H isotope fractionation on ΔG_r, that are observed in experiments.

We developed a bio-isotopic model, that relates the concentrations of intracellular metabolites in methanogens and ΔG_r to ¹³α, ²α and Δ₁₈ at a steady state. The model is based on a framework proposed for sulfate-reducing bacteria [4], and considers the full metabolic pathway, including electron carrier cycling, sodium pumping, and incorporation of H atoms from both H₂O and H₂. We calculated the theoretical equilibrium fractionations associated with each step of the metabolic pathway, and used the model to predict the reversibility degree of these steps at different conditions, to ultimately get the isotopic composition of CH₄ as a function of ΔG_r.

Our model elucidates the intracellular controls over stable isotope fractionation in methanogenesis. We find that as ΔG_r becomes increasingly negative, the enzymatically catalyzed reactions depart from equilibrium differentially. Departure from equilibrium of a pathway step inhibits expression of isotope fractionation of downstream steps, and shapes the observed net fractionation during methanogenesis. The model successfully predicts ¹³α- and ²α-ΔG_r relationships observed in lab cultures. In addition, it predicts Δ₁₈-²α-ΔG_r relationships, and identifies the key metabolic steps generating Δ₁₈ disequilibria.

[1] Valentine *et al.*, 2004, *GCA*. **68**, 1571–1590. [2] Okumura *et al.*, 2016, *PEPS*. **3**, 14. [3] Wang *et al.*, 2015, *Science*. **348**, 428–431. [4] Wing & Halevy, 2014, *PNAS*. **111**, 18116–25.