Experimental Study of Methane Isotopologue Fractionation During Microbial Methanogenesis

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The advent of CH_4 clumped isotopologue (e.g., $^{13}CH_3D$) analysis has enabled a more comprehensive investigation on the isotopic composition of microbial CH_4 . Laboratory cultures produce CH_4 with strong kinetic signals in the $^{13}CH_3D$ abundance, whereas environmental samples carry much less kinetic signals [1,2]. Previous studies have suggested that enzyme-mediated metabolic processes and environmental factors such as pH_2 can affect isotope fractionation during microbial methanogenesis [1,3]. However, it is unclear how fractionation mechanisms in bulk isotope and clumped isotopologue systematics are correlated.

We performed a series of culturing experiments to trace the origin of kinetic isotope effects during hydrogenotrophic methanogenesis. A hyperthermophilic methanogen *Methanocaldococcus bathoardescens* was grown in a fed-batch flow reactor. The isotopologue ratios among ¹²CH₄, ¹³CH₄, ¹²CH₃D and ¹³CH₃D were measured by tunable laser direct absorption spectroscopy, and the bulk isotope ratios of CO₂, H₂ and H₂O were measured by isotope ratio mass spectrometry.

Our results confirm the previous observation of metabolic state-dependent patterns in ${}^{13}C/{}^{12}C$ fractionation, approaching thermodynamic equilibrium during stationary phase [4]. D/H fractionation, however, shows convergence to the same degree of disequilibrium during stationary phase, apparently independent of temperature and pH_2 . $\Delta^{13}CH_3D$ values (the deviation of ${}^{13}CH_3D$ abundance from the expected statistical value) carry varying degrees of kinetic signals under all tested conditions, including a pH_2 range conceivable in the natural habitat of this organism [5].

The observed decoupling among ¹³CH₄, ¹²CH₃D and ¹³CH₃D fractionations can be used to locate the isotope sensitive and rate-limiting steps in the hydrogenotrophic methanogenesis pathway. This model can be used to correlate various environmental parameters with isotopic signatures observed in nature.

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68, 1571-1590. [4] Botz et al. (1996) Org. Geochem. 25, 255-262. [5] Ver Eecke et al. (2012) PNAS 109, 13674-13679.