

## Elemental mercury production in seawater by coastal bacterial assemblages

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Elemental mercury ( $\text{Hg}^0$ ) evasion from surface seawater plays a principal role in the marine mercury cycle.  $\text{Hg}^0$  production in seawater is mainly controlled by abiotic (photochemical) and biotic (microbial) processes. In this study, we established a method using the gamma-emitting radionuclide  $\text{Hg}$ -203 as a tracer to evaluate the transformation of  $\text{Hg}^{2+}$  and methylmercury (MeHg) to  $\text{Hg}^0$  in seawater by coastal bacterial assemblages. The method, which used traps containing gold-coated beads to capture  $\text{Hg}^0$  released into air from seawater, can provide rapid and reliable  $\text{Hg}^0$  measurements that avoid potential contamination. Several natural bacterial assemblages in surface seawater collected 8 km off Southampton, New York, were used in our experiments. These bacterioplankton were contained in seawater collected for years prior to our study and stored in the dark at 4°C. Remarkably, these bacteria were still able to rapidly produce  $\text{Hg}^0$  following picomolar additions of  $^{203}\text{Hg}^{2+}$  or  $\text{Me}^{203}\text{Hg}$  when brought up to 23°C in 2 days. Our results show that  $\text{Hg}^0$  production rates were independent of dissolved  $\text{Hg}^{2+}$  and MeHg concentrations, and were directly a function of bacterioplankton densities. Addition of antibiotics reduced  $\text{Hg}^0$  evasion to undetectable levels. These Hg evasion experiments showed that for 1  $\mu\text{m}$ -filtered Long Island coastal waters from the Atlantic with natural bacterial assemblages and bacterial densities of about  $1 \times 10^6 \text{ ml}^{-1}$ , approximately 25% of  $\text{Hg}^{2+}$  and 18% of MeHg were transformed to  $\text{Hg}^0$  in 4 days at ~23°C. In Long Island Sound waters, with  $5 \times 10^6$  bacterial cells  $\text{ml}^{-1}$ , 60% of  $\text{Hg}^{2+}$  and 19% of MeHg were converted to  $\text{Hg}^0$  and trapped in the air within 4 days. When bacterial assemblages were exposed to  $\text{Hg}^{2+}$ , the  $\text{Hg}^0$  production rate declined after one day, but the rate of  $\text{Hg}^0$  evasion from bacterial assemblages exposed to MeHg remained constant over 4 days, suggesting two distinct production pathways. Total  $\text{Hg}^0$  production for both  $\text{Hg}^{2+}$  and MeHg exposures at ~23°C were 6 times those at 4°C, indicating such transformations were mainly driven by metabolic processes.