

## Rhizosphere Control of Soil Carbon Association with Fresh Minerals

RACHEL NEURATH<sup>1</sup>, THEA WHITMAN<sup>2</sup>, PETER NICO<sup>3</sup>, JENNIFER PETT-RIDGE<sup>4</sup>, JIZHONG ZHOU<sup>5</sup>, ANDREW LIPTON<sup>6</sup>, PETER WEBER<sup>7</sup>, MARY FIRESTONE<sup>8</sup>

<sup>1</sup>UC Berkeley, Berkeley, CA rneurath@berkeley.edu

<sup>2</sup>University of Wisconsin, Madison, WI  
twhitman@wisc.edu

<sup>3</sup>LBNL, Berkeley, CA psnico@lbl.gov

<sup>4</sup>LLNL, Livermore, CA pettridge2@llnl.gov,

<sup>5</sup>University of Oklahoma, Norman, OK  
jzhou@rccc.ou.edu

<sup>6</sup>EMSL, PNNL, Richland, WA as.lipton@pnnl.gov

<sup>7</sup>LLNL, Livermore, CA weber21@llnl.gov

<sup>8</sup>UC Berkeley, Berkeley, CA mkfstone@berkeley.edu

The rhizosphere - the nexus of plant-soil-microbe interactions - is the largest active terrestrial C reservoir. As roots transfer organic compounds to the soil, the fate of this SOM is determined by (i) *who* is there (which microbial taxa), (ii) *what* chemical form the C is in, and (iii) *where* C is associated within the soil physical environment. We aim to develop a mechanistic understanding of plant-derived C association with soil minerals, investigating how SOM is protected from microbial degradation during growth of *Avena barbata*, a Mediterranean annual grass. We grew *A. barbata* with 99 atom% <sup>13</sup>C<sub>2</sub>O<sub>2</sub> and tracked <sup>13</sup>C-labeled photosynthates into soil microcosms where three mineral types were incubated: FeO-coated quartz, kaolinite, and ferrihydrite, representing a spectrum of reactivity and surface area. Mineral samples were collected at four timepoints during plant growth, ending at senescence. In the second phase of our experiment, <sup>13</sup>C-labeled roots from the initial study were ground and mixed with soil to represent root litter created during the long period of Mediterranean dry-season senescence. Again, the three mineral types were incubated in soil, with CO<sub>2</sub> collected throughout the incubation. Mineral microbial communities and C associations were characterized using bacterial and archaeal 16S and fungal ITS Illumina sequencing, total C and <sup>13</sup>C, <sup>13</sup>C-NMR, and FTIR. To investigate C-mineral associations at the scale of individual soil microorganisms, we used combined STXM and NanoSIMS molecular/isotopic imaging to measure the distribution of specific C functional groups and <sup>13</sup>C enrichment on the mineral surfaces. Our findings suggest that (1) while mineral reactivity enhances SOM association, the presence of even relatively non-reactive surfaces allows for SOM accumulation, (2) growing roots host fungal partners which directly shunt C to minerals, and (3) microbial colonization of fresh minerals differs depending on mineralogy.