

Physiological and ecological constraints on TEX₈₆ and GDGT provenance revealed by pure culture experiments and quinone biomarkers

FELIX J. ELLING^{1*}, KEVIN W. BECKER¹,
MARTIN KÖNNEKE¹, JAN M. SCHRÖDER¹,
SARAH J. HURLEY³, MARC MUEBMAN², ANN PEARSON³
AND KAI-UWE HINRICHS¹

¹MARUM, University of Bremen, 28359 Bremen, Germany

²Max-Planck-Institute for Marine Microbiology, 28359 Bremen, Germany

³Harvard University, Cambridge, MA 02138, USA

(*correspondence: felling@marum.de)

Archaeal glycerol dibiphytanyl glycerol tetraether (GDGT) membrane lipids are ubiquitous in the environment. They are used both as biomarkers for the abundance of living archaea and in the reconstruction of past sea surface temperatures using the TEX₈₆ proxy. However, widely observed discrepancies between *in situ* and TEX₈₆ temperatures in the marine water column indicate that the physiological and ecological controls on lipid composition in planktonic archaea are still poorly understood. Moreover, the relative contributions of *Eury-* and *Thaumarchaeota* to the total GDGT signal remain controversial.

Here, we demonstrate that the responses of membrane lipid compositions, and resulting TEX₈₆ values, to growth temperature strongly diverge in three closely related thaumarchaeal pure cultures, including *Nitrosopumilus maritimus* and two novel isolates from South Atlantic surface water. Moreover we show that GDGT composition in *N. maritimus* depends on growth stage, growth rate, and pH, but not on salinity. This indicates that the TEX₈₆ paleotemperature proxy is not solely dependent on temperature, but amalgamates several physiological and environmental factors such as phylogenetic composition and metabolic state of marine archaeal communities.

In order to identify diagnostic lipid biomarkers for the activity and abundance of specific archaeal clades, we studied the lipidomes of >30 cultivated representatives of the three major archaeal phyla *Eury-*, *Cren-*, and *Thaumarchaeota* using novel, comprehensive analytical protocols. We identified respiratory quinones that are involved in cellular energy transfer in *Thaumarchaeota*, as well as the unique membrane lipid methoxy archaeol. We demonstrate that these novel biomarkers are well suited for tracing the presence and activity of planktonic *Thaumarchaeota* independently from GDGTs, thus offering high potential for distinguishing thaumarchaeal and euryarchaeal sources of GDGTs.