

Palaeozoic biosphere and climate: Modes of marine primary production and methane cycling feedbacks

MEGAN ROHRSSSEN¹, GORDON D. LOVE*¹
AND CHRISTOPHER T. REINHARD²

¹Department of Earth Sciences, University of California,
Riverside, CA, 92521, USA; *Correspondence:
glove@ucr.edu

²Department of Geological and Planetary Sciences, California
Institute of Technology, Pasadena, CA, 91125, USA

Lipid biomarker records of microbial community structure during the Late Ordovician through Early Silurian (~449-439 Ma) indicate substantial differences in the balance of bacterial versus algal primary production through the Late Ordovician, largely in response to climatic change associated with the Hirnantian glaciation ([1]). Our biomarker records suggest that these warm Palaeozoic intervals were associated not only with elevated atmospheric $p\text{CO}_2$ (~4-16x preindustrial atmospheric levels, PAL), but also with a higher average flux of oceanic methane, exerting a potential positive feedback on hothouse climates in the Paleozoic. To refine the assessment of a methanotrophic contribution to lipid biomarkers, we have conducted compound-specific carbon isotope analyses on hydrocarbon extracts from Anticosti Island (Canada) the Vinini Formation (USA), the Maquoketa Formation (USA), and OM9 drill core (Estonia).

Recent models have demonstrated that a relatively modest increase of 4-5x preindustrial $p\text{CH}_4$ can generate 2-3°C of warming [2]. Methane cycling feedbacks on climate must have been of similar or greater impact in the Palaeozoic, a period of warm conditions and reduced oxidant availability [3] which may have driven a larger proportion of remineralized organic matter through microbial methanogenesis and attenuated the efficacy of combined anaerobic methane oxidation processes as a throttle for methane fluxes to the atmosphere. Because the loss rate of atmospheric methane scales inversely with atmospheric methane abundance [4], increased fluxes to the atmosphere will often translate into higher steady-state concentrations. We explore the possible effects of this for Palaeozoic times with a series of simple mass balance calculations given a range of carbon flux and anaerobic remineralization scenarios.

[1] Rohrsen, *et al.* (2013), *Geology* 47, 127-130. [2] Beerling, *et al.* (2011), *PNAS* 108, 9770-9775. [3] Luo, *et al.* (2010), *EPSL* 300, 101-111. [4] Schmidt & Shindell (2003), *Paleoceanography* 18, 1-9.

Solubility and TRLFS studies on Nd(III)/Cm(III) complexation with gluconate in NaCl and CaCl₂ media.

H.ROJO^{1,2}, X. GAONA¹, TH. RABUNG¹, M. GARCIA³,
T. MISSANA³ AND M. ALTMAIER¹

¹Institute for Nuclear Waste Disposal, Karlsruhe Institute of
Technology, Karlsruhe, Germany (henar.rojo@kit.edu)

²Laboratory for Waste Management, Paul Scherrer Institut,
Villigen PSI, Switzerland

³CIEMAT, Research Centre for Energy, Environment and
Technology, Madrid, Spain

Radionuclide sorption and solubility in cementitious systems can be affected by the presence of organic ligands. Gluconic acid (GLU) is a poly-hydroxocarboxylic acid expected in repositories for low and intermediate-level radioactive waste as a component of cementitious materials. The formation of very stable An(III)-GLU complexes has been reported in the literature, although in contrast to An(IV) no ternary species Ca-An(III)-GLU have been described so far. These species may play a relevant role in cementitious and saline environments, where high Ca^{2+} concentrations are expected in certain scenarios.

Undersaturation solubility studies with $\text{Nd}(\text{OH})_3(\text{am})$ were conducted in inert gas (Ar) gloveboxes at $22 \pm 2^\circ\text{C}$. Samples were prepared in dilute NaCl (0.1 M) and CaCl_2 (0.1 and 0.25 M) solutions as background electrolytes. Parallel experimental series were prepared with $\text{pH}_c = \text{constant} \sim 12$ and $10^{-6} \leq [\text{GLU}] \leq 10^{-2}$ M, and with $[\text{GLU}] = \text{constant} = 10^{-3}$ M and $9 \leq \text{pH}_c \leq 13$. TRLFS measurements were performed with $\sim 10^{-7}$ Cm(III) per sample, with NaCl (0.1 M) and CaCl_2 (0.1 and 0.25 M) as background electrolytes. In the NaCl systems, three different concentration levels of Ca^{2+} were considered: 0, 10^{-3} M and 10^{-2} M. The initial concentration of GLU in all samples (10^{-6} M) was increased to $3 \cdot 10^{-3}$ M by step-wise additions of NaCl-NaGLU or CaCl_2 -CaGLU₂ solutions of appropriate ionic strength.

The solubility of $\text{Nd}(\text{OH})_3(\text{am})$ remains unaffected by GLU in 0.1 M NaCl solutions. On the other hand, solubility of $\text{Nd}(\text{OH})_3(\text{am})$ in 0.1 and 0.25 M CaCl_2 solutions is clearly increased by GLU under hyperalkaline conditions. The species forming are pH-dependent and unequivocally involve the participation of Ca^{2+} , with the likely formation of a Nd-GLU complex with stoichiometry 1:2. No further increase of Nd(III) concentration is observed at $[\text{GLU}]_{\text{tot}} \geq 10^{-3}$ M, resulting in an upper solubility limit at $[\text{Nd}] \sim 10^{-6.5}$ M which suggest the formation of a new Ca-Nd-GLU solubility limiting solid phase. Consistently with Nd(III) solubility data, TRLFS confirms the key role of Ca^{2+} in the complexation process, with the likely formation of a Ca-Cm(III)-GLU complex with Ca:GLU ratio 1:1. Two Ca-Cm-GLU species are further identified in 0.1 M and 0.25 M CaCl_2 solutions.