REE behavior during the formation of Sn-W deposits

J.A. POPOVA^{1*}, A.YU. BYCHKOV¹, S.YU. NEKRASOV¹ AND T. SUSCHEVSKAJA²

¹Moscow State Universiy, Moscow, Russia (*correspondence: julka_p@rambler.ru)
²GEOKHI RAS, Moscow, Russia

REE behaviour was considered for two Sn-W deposits,

Svetloe and Iul'tin (Chukotka, Russia). Both deposits take place within the Iul'tin ore district. The isotopic composition of oxygen and hydrogen allowed to conclude that meteoric waters were probable source of hydrothermal fluids in these deposits [1]. REE content in fluorites may be used as an indicator to determine the nature and the evolution of fluids [2]. Regularities of REE spectra changes indicate the presence of two generations of fluorite. REE concentations in thehydrothermal solutions in equilibrium with fluorites were obtained by conversion of two generations of fluorite into fluids using the distribution coefficients of mineral / fluid [2]. For the first fluorite generation the fluid is magmatic one, for the second fluorite generation the fluid is exogenous. Mixing these two fluids leads to the deposition of ore. The deposition of cassiterite and arsenopyrite occurs when acidic magmatic fluid carrying the ore elements mixed with alkaline meteoric water. As a result pH increases and the following association is formed:

 $3Sn^{II}Cl_2^{\circ}+2As^{III}(OH)_3^{\circ}+2FeCl_2^{\circ}+2H_2S^{\circ}=$

 $=3Sn^{IV}O_2+2FeAs^{\circ}S+10HCl^{\circ}$. The wolframite deposition occurs under the following reaction:

NaHWO₄°+FeCl₂°=FeWO₄+Na⁺+2Cl⁻+H⁺.

The compositon of wolframite changes in the vertical section of Iul'tin deposite from 47% to 64% mol% FeWO4. For Svetloe deposite it is shown that wolframite is depleted of iron (from 39 to 5 mol% FeWO4) under the joint deposition of wolframite and arsenopyrite.

[1] Sushchevskaya T. *et al.* (2000) *Geochem. Int.* **38**. Suppl. 2 . P. 123. [2] Raimbault L. (1985) *Bull. Mineral* **108**. P.737.

Methylotrophy in Yellowstone National Park hot springs

A.T. PORET-PETERSON^{1*}, S.J. ROMANIELLO¹, N. ZOLOTOVA¹, Z. MARTINEZ¹, J.J. ELSER² AND A.D. ANBAR^{1,3}

¹School of Earth and Space Exploration, Arizona State University, PO Box 871404, Tempe, AZ 85287, USA (*correspondence: aporetpe@asu.edu)

²School of Life Sciences, Arizona State University, PO Box 874501, Tempe, AZ 85287, USA

³Department of Chemistry and Biochemistry, Arizona State University, PO Box 871604, Tempe, AZ 85287, USA

Autotrophic microorganisms in hot springs, like their mesophilic counterparts in other aquatic ecosystesm, likely play a major role in supplying reduced organic C to these systems [1]. However, hot springs are surface expressions of geologic activity that releases compounds that may serve equally well as sources of C, such methane (CH₄). Indeed, CH₄ assimilation pathways may be less energetically expensive in terms of ATP and reductant than the Calvin Cycle or alternative pathways for CO₂ fixation [2]. Thermodynamic calculations also show that microbes capable of aerobic CH₄ oxidation can thrive in hydrothermal ecosystems due to the amount of energy released from its catabolism [3]. Several recent studies have isolated CH₄oxidizers from geothermal environemnts [4].

The goal of this study was to investigate whether methylotrophs (utilizers of reduced C1 compounds) were present and active in various thermal features in Yellowstone National Park. To accomplish this objective, we incubated hot spring sediments and microbial mats in microcosms amended with stable-isotope labeled and unlabeled methanol (CH₃OH) overnight at in situ temperatures. We found evidence for methylotrophy in the sediments of slightly acidic hot springs (pH 6.0) based on ¹³C enrichment of CO₂ and bulk sediment. To complement our isotopic data, we extracted DNA from hot spring sediments and retrieved functional genes encoding key enzymes in aerobic CH₄ and CH₃OH oxidation. Sequencing of these genes suggests that a complex methylotrophic community may be present in the hot spring, as clones grouped with gammaproteobacterial obli-gate methanotrophs and betaproteobacterial obligate methylo-trophs.

Azam & Malfatti (2007) *Nature* 5, 782–791. [2] Berg (2011) *Appl. Environ. Microbiol.* 77, 1925–1936. [3] Shock *et al.* (2010) *Geochim. Cosmochim. Ac.* 17, 4005–4043.
 Trotsenko *et al.* (2009) *Microbiol.* 78, 387–401.

Mineralogical Magazine